A.C. Gouda, F.R. Hay, Y.D. Moukoumbi, G.L. Djedatin and M.N. Ndjiondjop (2025). *Seed Science and Technology*, **53**, 1, 105-119. https://doi.org/10.15258/sst.2025.53.1.09



Development of an efficient seed dormancy breaking method for African rice (*Oryza glaberrima*)

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(Submitted June 2024; Accepted March 2025; Published online April 2025) (Handling Editor: A.A. Powell)

Abstract

Seed dormancy is a significant factor limiting the conservation, evaluation, distribution and utilisation of African cultivated rice, *Oryza glaberrima*. In this study, 20 accessions from the AfricaRice genebank were grown at the AfricaRice Mbé station in Côte d'Ivoire, harvested manually, dried at 15% RH and 15°C, and stored at -20°C for one month before use. Seeds underwent dry heat treatment at 50 or 60°C for 2, 4, 7, 14, 21 or 28 days, or were treated with: 0.5, 1, 1.5 or 2 M KCl; 0.022, 0.112, 0.224 or 0.448 M HNO₃; 0.05, 0.1, 0.2, 0.4, or 0.5 M KH₂PO₄; and 2.89, 5.77 or 14.44 mM gibberellic acid (GA₃). Most treatments on intact seeds did not significantly increase germination, except for those treated at 50°C for 21 or 28 days. Dehulled seeds germinated effectively with all treatments except HNO₃ solutions. After validating the treatment on 287 *O. glaberrima* accessions, it was concluded that dry heat treatment at 50°C for 28 days effectively breaks dormancy. This protocol improves germination percentages of *O. glaberrima*, enhancing its use in various studies and aiding better decision-making in the management of *O. glaberrima* accessions in genebanks.

Keywords: African rice, dormancy, dry after-ripening, germination, gibberellic acid, heat, Oryza glaberrima

Introduction

Rice (*Oryza* spp.) is a staple food for more than half of the world's population (Pandey *et al.*, 2010; Futakuchi *et al.*, 2021). It is the third cereal crop in terms of production, after maize and wheat, with a world production of 518.2 million tonnes in 2021 (FAO, 2021). The African continent has the second largest rice consumption, after Asia (Saito *et al.*, 2015; Futakuchi *et al.*, 2021). In Africa, the share of rice in total cereal production has steadily increased from 9.3 to 17.8% between 1961 and 2018 (FAOSTAT, 2020). Paddy rice production in Sub-Saharan Africa was estimated at 26.5 million tons from a total of 11.95 million hectares of harvested area in 2018 (Ndindeng *et al.*, 2021).

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In West Africa, rice is a strategic commodity for food security. Rice consumption quickly expanded from the 1960s, driven by demographic growth, urbanisation and changes in consumer preferences (Mendez del Villar and Lançon, 2015; Sekiya *et al.*, 2020). The annual per capita rice consumption steadily increased from 10 kg in 1961 to 54 kg in 2017 (Osinski and Sylla, 2018). Annual rice consumption in Africa, per capita is projected to increase by about 5 kg over the period 2019–2028 (OECD/FAO, 2020). However, only 60% of consumer demand is met through local production and the rest is imported (Zenna *et al.*, 2017). In the 2020/2021 trade year, Africa imported around 16.6 million metric tons of rice (Statista, 2023).

Although considerable efforts are increasingly being made to increase rice productivity worldwide, through the introduction of high-yielding varieties and the adoption of new cultivation practices, rice production in Africa is still subject to several biotic and abiotic constraints. Biotic stress is caused by bacteria, fungi, viruses, pathogenic nematodes, insect pests, weeds and invasion of parasitic plants (Ye *et al.*, 2019). Regarding abiotic stress, drought remains one of the major factors that hinder plant growth or development (Nelson *et al.*, 2014; Pandey and Shukla, 2015). Other factors such as soil salinity, acidity, iron and alumina toxicity, and phosphorus and zinc deficiency are all abiotic factors that hamper rice production (Sikirou *et al.*, 2015; Fahimirad and Ghorbanpour, 2018; Pathak *et al.*, 2021).

Seed dormancy can be defined as an innate property of the seed that prevents germination even under suitable environmental conditions (Finch-Savage and Leubner-Metzger, 2006). Seed dormancy is a complex trait, affected by multiple genes and environmental factors (Graeber *et al.*, 2014; Lu *et al.*, 2018), and an important component of plant adaptation (Donohue *et al.*, 2005; Huang *et al.*, 2010). It is regulated by the antagonistic hormones, abscisic acid (ABA) and gibberellic acid (GA) (Finkelstein *et al.*, 2008; Shu *et al.*, 2016; Née *et al.*, 2017a, b). Seed dormancy can be considered an undesirable trait by farmers, affecting crop establishment. Hence, selection to reduce levels of seed dormancy occurred during the rice domestication process and most cultivated plants germinate uniformly and rapidly after sowing, contrary to their wild ancestors (high dormant varieties/germplasms/accessions) (Kilian *et al.*, 2009). On the other hand, too low dormancy levels reduce seed quality for sowing and trigger pre-harvest germination, resulting in yield losses in cereals (Gubler *et al.*, 2005).

Seed dormancy in *Oryza* exhibits significant variation depending on the species and subspecies, reflecting the evolutionary and ecological adaptations of the genus. For example, differences in dormancy between different variety groups of *O. sativa* L. (Zhang *et al.*, 2020) highlight the influence of domestication and selection on this trait. Furthermore, the identification of over 150 quantitative trait loci (QTLs) associated with seed dormancy in *O. sativa* underscores the complexity of its genetic regulation (Zhang *et al.*, 2020). Comparative studies reveal that wild species such as *O. rufipogon* Griff. exhibit higher dormancy levels than cultivated species like *O. sativa* and *O. glaberrima* Steud. (Veasey *et al.* 2004). Interestingly, *O. glaberrima* displays stronger dormancy than *O. sativa*, as confirmed by Veasey *et al.* (2004) and Doku *et al.* (2016), possibly reflecting its adaptation to African environments and its role in traditional farming systems. These findings highlight the need for effective dormancy-breaking treatments tailored to the specific characteristics of *O. glaberrima*. The Africa Rice Center (AfricaRice) genebank conserves 20,681 registered rice samples, representing five African indigenous wild species (*O. barthii* A.Chev., *O. longistaminata* A.Chev & Roehr., *O. eichingeri* Peter, *O. punctata* Kotschy ex Steud. and *O. brachyantha* A.Chev & Roehr.), as well as both modern and traditional *O. glaberrima* and *O. sativa* varieties (Ndjiondjop *et al.*, 2017). *O. glaberrima* accounts for approximately 17% (> 3,000 accessions) of the collection (Ndjiondjop *et al.*, 2022). An important process that is part of the genebank's routine operations is seed viability testing, usually through a germination test. To ensure that the germination results reflect the actual viability, it is essential that an effective dormancy breaking treatment is applied. For rice, different seed dormancy breaking treatments have been proposed, for example, removing the seed hull (palea and lemma), varying temperature, giving seed a dry heat treatment or applying chemicals (Doku *et al.*, 2016; Shiratsuchi *et al.*, 2017).

The AfricaRice genebank team have been placing *O. glaberrima* seeds at 50°C for five days to break dormancy. This allowed categorisation of the seed lots as: (*i*) weakly dormant; (*ii*) moderately dormant; and (*iii*) strongly dormant. However, this approach proved insufficient for consistently breaking dormancy across all accessions, as it did not result in high germination for many seeds. This limitation could be due to the variation in dormancy levels among accessions, which might require a more robust treatment to effectively break dormancy and achieve higher germination. The aim of the present study was to develop a more efficient treatment for routine use in viability testing *O. glaberrima* accessions. Specifically, the objective of this study was to (*i*) evaluate the extent of seed dormancy and its response to a variety of dormancy breaking treatments on 20 randomly selected *O. glaberrima* rice accessions (both intact and dehulled seeds); (*ii*) validate the best dormancy breaking treatment method on 287 accessions, representing the *O. glaberrima* mini-core collection.

Materials and methods

Fresh rice seed production, drying, packaging and conservation at -20°C.

Samples of seeds from 20 randomly selected accessions and 287 mini-core accessions of African rice (*Oryza glaberrima*) were taken from the AfricaRice genebank (table 1). Seeds were sown for multiplication in the field in May 2021 (irrigated lowland ecology) according to a randomised incomplete block design (RIBD) without repetition. Irrigation, weed management, fungicide/insecticide application, fertilizer application, rouging to eliminate off-types and other routine field cultural procedures were all undertaken as described in the rice knowledge bank website (http://www.knowledgebank.irri.org/step-by-step-production). Seeds were harvested when 80–85% reached maturity and seed moisture content was $18 \pm 1\%$ (determined using a Rice Flour Moisture Tester, SATAKE model MOISTEX SS7). Then, seeds were dried in the sun (August 2021), under a maximum temperature of 28.2°C and a minimum temperature of 21.7°C, to decrease their moisture content from 18 to $13 \pm 1\%$. After threshing, cleaning, grading and labelling, seeds were dried further in a drying room where the conditions are controlled (15°C and 15% RH).

For the 20 random accessions, samples were packed in aluminium foil pouches according to the number of dormancy-breaking treatments to be tested. Seeds of the remaining accessions were also sealed inside aluminium foil pouches. Following Africa-Rice standard procedures, all the packages were stored at -20°C, in the long-term storage room until one month before use. Thereafter, packages were allowed to equilibrate at 2-5°C for approximately 48 hours, and then at room temperature $(23 \pm 2^{\circ}C)$ before opening.

Seed viability and dormancy evaluation

Before proceeding with dormancy-breaking treatments, the viability of seeds from the 20 accessions was determined based on a tetrazolium test (ISTA, 2021). For each accession, 100 seeds were preconditioned by soaking in distilled water at room temperature $(23 \pm 2^{\circ}C)$ for 18 hours and then, longitudinally dissected through the embryo and $\frac{3}{4}$ of the endosperm before being soaked in 1% tetrazolium solution for two hours at 25°C in the dark. After that, seeds were washed twice with distilled water. Seeds were scored as viable when the embryo was completely stained. Viability percentage of each accession was estimated based on the total number of seeds with the embryo completely stained divided by the total number of seeds tested.

Dormancy was assessed for all 307 accessions by germination of intact seeds in 90 mmdiameter Petri dishes, on filter paper moistened with distilled water, at room temperature $(23 \pm 2^{\circ}C)$. Each accession was represented by three random samples of 100 seeds and germination was monitored daily over a period of 14 days. As also adopted by Shi *et al.* (2021), germination was considered here as emergence of the roots (root length ≥ 2.0 cm). Dormancy percentage of each accession was computed based on the total number of seeds not germinated divided by the total number of seeds sown. Seed lots with germination $\leq 25\%$ were considered to be strongly dormant, whereas those with seed germination $\geq 50\%$ were considered to be weakly dormant (Waheed *et al.*, 2012).

Dormancy-breaking treatments

Intact and dehulled seeds from each of the 20 accessions were used to test the effect of the different treatments described below. Three random samples, each consisting of 20 dehulled seeds and 20 intact seeds, were used for all treatments. Seeds of each accession were geminated as above and the number of intact and dehulled seeds germinated for each accession counted.

Heat treatments

Samples (intact and dehulled seeds from each accession) were incubated (dry) at 50°C or at 60°C, for 2, 4, 7, 14, 21 and 28 days in an oven (BINDER Machin). After that, seeds packages were allowed to equilibrate at room temperature $(23 \pm 2^{\circ}C)$ during 12 hours before use. Both dehulled and intact seeds were germinated in Petri dishes under previous germination conditions described above.

Chemical treatments

Four chemical treatments at different concentrations were applied to intact and dehulled seeds: (*i*) 0.5, 1, 1.5 or 2 M KCl; (*ii*) 0.022, 0.112, 0.224 or 0.448 M HNO₃; (*iii*) 0.05, 0.1, 0.2, 0.4 or 0.5 M KH₂PO₄; (*iv*) 2.89, 5.77 or 14.44 mM gibberellic acid (GA₃). For each chemical and concentration tested, seeds were soaked for 24 hours and then washed twice with distilled water before germination in Petri dishes as described above.

Validation of the best treatment or protocol

Three random samples of 100 seeds from each of the 287 accessions collected from the *O*. *glaberrima* minicore set were treated with the best treatment identified from all previously tested treatments. Seeds were germinated in Petri dishes under the same germination conditions described above.

Statistical analysis

Data collected during the different treatments tested on the 20 accessions were imported into R program v4.0.3 (The R Development Core Team, 2011) and then visualised using the *ggplot2* R package version 3.3.5 (Ginestet, 2011).

The germination data was coded into a binary format using the \geq 50% accepted germination threshold for *Oryza glaberrima* species as applied by dormant Waheed *et al.* (2012) in *Oryza* genus. Accessions × pre-treatments with germination \geq 50% were coded as 1 (successful germination) while accessions with < 50% germination were coded as 0 (unsuccessful germination). The data were then analysed in R software (version 4.4.0) using logistic regression with binomial error and logit function whereby the germination variable was fitted as the dependent variable with accessions and treatments as an independent variable (Olbana *et al.*, 2023). The predicted probability of a successful germination result for each accession is presented.

Results

Seed embryos for all *Oryza glaberrima* accessions seeds were stained red by 1% TZ solution, thus, all seeds were viable. For 16 of the 20 accessions, there was no germination, i.e. 100% dormancy. For accessions WAB0029199, WAB0029415, WAB0024666 and WAB0013353, dormancy percentages were 95, 95, 85 and 80%, respectively (table 1).

No germination was noted with intact and dehulled seeds not treated with the four chemical solutions. When intact seeds were treated with the different KH_2PO_4 solutions the germination percentage was less than 50% (figure 1). However, germination ranging from 18.25 to 57.25% for 0.5 and 0.05 M KH_2PO_4 , respectively, was noted for dehulled seeds (averaged across different accessions). No effective germination was observed with intact seeds treated with different HNO₃ concentrations expect for the 0.224 M concentration (mean germination 10.5%; figure 1). For dehulled seeds, 0.022 M HNO₃ solution resulted in a mean germination percentage of 32% but no germination was observed with other HNO₃ concentrations. Different KCl solutions barely had any effect on overcoming the

Accession number	Ecology	Biological status	Country of Origin	Dormancy (%)
WAB0029838	Rainfed Lowland	Traditional cultivar/landrace	Brazil	100
WAB0039053	Rainfed Lowland	Traditional cultivar/landrace	Burkina Faso	100
WAB0026193	Rainfed Lowland	Traditional cultivar/landrace	Nigeria	100
WAB0039052	Rainfed Lowland	Traditional cultivar/landrace	Burkina Faso	100
WAB0032322	Rainfed Lowland	Traditional cultivar/landrace	Nigeria	100
WAB0039066	Rainfed Lowland	Traditional cultivar/landrace	Burkina Faso	100
WAB0032311	Irrigated lowland	Traditional cultivar/landrace	Liberia	100
WAB0032309	Rainfed Lowland	Traditional cultivar/landrace	Liberia	100
WAB0011747	Shallow Forest Swamp	Traditional cultivar/landrace	Nigeria	100
WAB0029725	Rainfed Lowland	Traditional cultivar/landrace	Brazil	100
WAB0032272	Irrigated lowland	Traditional cultivar/landrace	Nigeria	100
WAB0001743	Upland	Traditional cultivar/landrace	Liberia	100
WAB0032570	Rainfed Lowland	Traditional cultivar/landrace	Brazil	100
WAB0032426	Irrigated lowland	Traditional cultivar/landrace	Liberia	100
WAB0029199	Upland	Traditional cultivar/landrace	Côte d'Ivoire	95
WAB0024666	Upland	Traditional cultivar/landrace	Côte d'Ivoire	85
WAB0029415	Irrigated lowland	Traditional cultivar/landrace	Guinea	95
WAB0013353	Irrigated lowland	Traditional cultivar/landrace	Guinea	80
WAB0032315	Rainfed Lowland	Traditional cultivar/landrace	Liberia	100
WAB0037616	Upland	Traditional cultivar/landrace	Senegal	100

Table 1. Passport information for the 20 accessions of *Oryza glaberrima* used for the dormancy-breaking experiments.



Figure 1. Germination of intact and dehulled seeds of 20 *Oryza glaberrima* rice accessions. Seeds were treated with different concentrations of KH_2PO_4 , HNO_3 , KCl or GA_3 . The box-and-whisker plots displayed the distribution of germination percentage for the 20 accessions tested with each chemical treatment.

dormancy of intact seeds (figure 1), but when dehulled seeds were treated with the different KCl solutions, mean germination was 68, 66.50, 57.50 and 43% for 0.5, 1, 1.5 and 2 M KCl, respectively. Very low germination was observed for intact seeds treated with GA₃ (germination percentage \leq 50%; figure 1) but GA₃ was effective in breaking dormancy of dehulled seeds. The mean germination values were 89.75, 93.75 and 94.25% for 2.89, 5.77 or 14.44 mM GA₃, respectively.

Intact seeds treated at 50 or 60°C for at least seven days showed some germination, but high levels of dormancy remained (figure 2). Treating intact seeds at 50°C for 14 days appeared to be more effective than treating at 60°C for the same length of time (64.75 vs. 52%). When intact seeds were heated at 50°C for more than 14 days, mean germination increased, e.g. from 86.75 to 90.5% after heating for 21 or 28 days, respectively.



Figure 2. Germination percentages of intact and dehulled seeds evaluated for 20 rice accessions belonging to *Oryza glaberrima* (Steud). Intact seeds from each accession were placed at 50°C for 2, 4, 7, 14, 21 or 28 days, or at 60°C for 2, 4, 7 or 14 days; dehulled seeds were placed at 50°C for 2, 4, 7 or 14 days, or at 60°C for 2, 4, 7 or 14 days. The box-and-whisker plots show the distribution of germination percentage for the 20 accessions tested with each heat treatment.

Germination probability value was 0 for all 20 accessions (intact seeds) treated with all chemical solutions at different concentrations, except for two accessions (WAB0024666 and WAB0013353, values ≈ 0.52). Also, these two accessions showed a probability of $\geq 50\%$ germination with dry heat treatment at 60°C for 14 days (values \approx 1). For intact seeds dried using heat at 50°C for 28 days, the predicted marginal means for the probability of $\geq 50\%$ germination were 1 for the 20 accessions. However, germination probability values were higher (≥ 0.85) for almost all dehulled seeds treated with various KCl and GA₃ concentrations, as well as with heat treatments at 50 and 60°C for 4 to 14 days (figure 3). Based on the results, it appears that heat treatment at 50°C for 28 days with intact seeds is effective in achieving $\geq 85\%$ germination for all 20 accessions (probability of 1).



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Figure 3. Effect of physical and chemical treatments on the mean germination probability of *Oryza glaberrima* seeds. Intact and dehulled seeds were subjected to various treatments, including dry heat treatment at 50°C or 60°C for 2 to 14 days, as well as chemical treatments with KCl, KH₂PO₄, HNO₃, and gibberellic acid (GA₃) at different concentrations. The mean germination probability was calculated for each treatment.

The mean germination of untreated seeds of 287 *O. glaberrima* accessions was 10.4% (figure 4A). More than 95% of the 287 accessions had less than 50% germination. After dry heat treatment at 50°C for 28 days, the overall mean germination for the 287 accessions was 90.01% (figure 4B).



Figure 4. Summary of mean germination percentages of intact seeds estimated for the 287 accessions of rice belonging to *Oryza glaberrima* species. Data was collected from three repetitions of each accession. (A) intact seeds not heated before the germination test; (B) intact seeds heated in an oven at 50°C for 28 days before the germination test.

Discussion

In rice genebanks such as AfricaRice and IRRI where *O. glaberrima* germination tests are routinely performed to monitor the viability of (*i*) rice accessions stored in mediumand long-term storage, (*ii*) rice accessions to be distributed and (*iii*) newly acquired seeds, seed dormancy can be a very disadvantageous factor. For rice genebanks such as AfricaRice, seed dormancy can be a disadvantageous factor during both routine monitoring of the viability of *O. glaberrima* accessions in the genebank and also testing of newly acquired seeds.

This could result in misleading viability data, which can lead to erroneous management decisions. In addition, accurate measurement of seed viability is very important for genebanks as they must ensure the distribution of viable seed. Poor germination due to an inadequate dormancy breaking treatment could also lead to early scheduling of multiplication and/or seed stock regeneration activities. Rice seed dormancy can be a problem for farmers because (*i*) it affects the rapid and uniform germination, (*ii*) vigour of seeds during the early growth stages of the crop, thus reducing their productivity. (Tung and Serrano, 2011; Waheed *et al.*, 2012). Previous studies exposed that dormancy is higher in fresh seeds than those stored over time (Ilyas and Diarni, 2007; Bewley and Nonogaki, 2017). Kumar *et al.*, (2009) demonstrated that dormancy period in Asian Basmati 370 rice could be around 50 days after harvest. In the present study, only new fresh seeds that supposed will have a very high dormancy level were used.

Our seed viability testing results, performed using a 1% TZ solution, showed that all *O. glaberrima* seeds tested were viable, but none of them germinated in the control germination tests. These results demonstrated high percentages of dormancy in *O. glaberrima* accessions, indicating the need for proper dormancy breaking methods. Our findings are consistent with those of Kuya *et al.* (2019) who also stated that African rice *O. glaberrima* had stronger dormancy than *O. sativa*. This led them to place their samples at 50°C for five days and store them at room temperature for about six months after harvest, with the aim of breaking their dormancy before use. According to Waheed *et al.* (2012), a species is considered highly dormant when seed dormancy percentage is greater or equal to 75%. Compared to our results, the seed lots of all 20 accessions have a high dormancy (dormancy percentage $\geq 75\%$).

There was no significant germination when intact seeds were treated with the different KH_2PO_4 solutions (germination percentage $\leq 50\%$). Similar results were also found by Ros *et al.* (2000) in intact seedlings of *O. sativa* rice 'IR66'. However, a different result to ours was obtained by Bam *et al.* (2006) who noted a germination percentage around 94% with *O. sativa* 'IDSA 85', when the seeds were treated with 0.5% (w/v) KH_2PO_4 solution for 24 hours as in our study.

No significant germination was noted with intact and dehulled seeds treated with different HNO₃ concentrations. Yuningsih and Wahyuni (2015) reported that some dormancy breaking methods recommended by the International Seed Testing Association (ISTA) were not so effective in breaking dormancy. One of these methods was 1 M HNO₃ solution; this method was ineffective to break dormancy in Indonesian rice varieties, consistent with our results. Concerning dehulled seeds soaked in different HNO3 concentration, we suspect that such concentrations ($> 0.1 \text{ M HNO}_3$ for 24 hours soaking) could be fatal to the embryo in dehulled seeds, which is why no seeds germinated. Such suspicions were also formulated by Yuningsih and Wahyuni (2015). Our results showed that different KCl solutions tested barely had any effect on overcoming the dormancy of intact seeds. Bam et al. (2006) were able to efficiently break dormancy in 'IDSA 85' (89% germination) by soaking seeds in 0.5% (w/v) KCL solution for 24 hours. These results confirm the hypothesis that O. glaberrima seed dormancy level is higher than for O. sativa. Similar results to ours were also obtained by Brooks et al. (2020) who tried to improve upland rice seed germination and vigour under salinity and water stresses using KCL, KNO₃ and MgCl₂ solutions.

Very low germination was observed with intact seeds treated with GA₃. Compared to results reported by Waheed *et al.* (2012), germination percentage computed on intact seeds of the cultivated variety 'Swat-1' (*O. sativa*) is higher than those found in our study with intact seeds treated with the three different concentrations of GA₃ (55.9% versus 4.24 to 6.25%). However, GA₃ was effective in breaking dormancy of dehulled seeds. The average percentage germination obtained with dehulled seeds treated with GA₃ at 14.44 mM are slightly similar to those published by Mutinda *et al.* (2017) on intact seeds of *O. sativa* 'Basmati 370' also treated for 24 hours with a GA₃ solution at 14.44 mM (94.25 versus 95.7%). Several studies have reported the effectiveness of GA₃ solution in breaking dormancy in various cereals by promoting germination (Shalinee *et al.*, 2019; Wu and Shen, 2021; Ahmed and El Dessougi, 2023; Nedunchezhiyan *et al.*, 2023).

METHOD TO BREAK SEED DORMANCY IN AFRICAN RICE

Intact *O. glaberrima* seeds heated at 50°C for 28 days showed high germination (90.5%) which is higher than those obtained by Hanumanthappa *et al.* (2016) and Waheed *et al.* (2012) on both wild and cultivated *O. sativa* accessions. Timple *et al.* (2018) also suggested that the dormancy of intact wild *Oryza* species seeds, including *O. barthii*, a wild rice species endemic to Africa and known as the progenitor of African cultivated rice, *O. glaberrima* (Bessho-Uehara *et al.*, 2017), can be effectively alleviated by dry heat treatment at 50°C and sowing directly in KNO₃, or by pre-soaking heat-treated seeds in GA₃ or H₂O₂ for 18 hours before soaking in distilled H₂O for a further 18 hours prior to sowing.

The mean germination of untreated seeds of 287 *O. glaberrima* accessions was very low confirming high levels of dormancy in fresh seeds of this mini-core germplasm (figure 4). According to the scale adopted by Waheed *et al.* (2012), the 287 accessions were grouped into three groups based on their dormancy level: (*i*) 254 accessions that were highly dormant (germination $\leq 25\%$); (*ii*) 20 accessions with medium dormancy (germination between 26 and 50%) and 13 accessions with low dormancy (germination $\geq 50\%$). This dormancy was effectively overcome by treating the seeds with dry heat treatment at 50°C for 28 days.

Conclusion

We strongly recommend that intact seeds of *Oryza glaberrima* be heated in an oven at 50°C for 28 days before sowing. For seeds with very high dormancy, heating at the same temperature for more than 28 days may yield better results. It provides a well-optimized protocol that can be used by rice genebanks, researchers and breeders, to efficiently break dormancy in *O. glaberrima* for its enhanced utilisation.

Acknowledgements

The present study was supported by a grant given to Marie Noelle Ndjiondjop (AfricaRice) BMZ Funding of Genebank Uplift from Deutsche Gesellschaft für International.

References

- Ahmed, T.M.M. and El Dessougi, H.I. (2023). Breaking seed dormancy in Argemone mexicana L. seeds. University of Khartoum Journal of Agricultural Sciences, 22, 95-111. <<u>https://doi.org/10.53332/uofkjas.</u> v22i1.1829>
- Bam, R.K., Kumaga, F.K., Ofori, K. and Asiedu, E.A. (2006). Germination, vigour and dehydrogenase activity of naturally aged rice (*Oryza sativa* L.) seeds soaked in potassium and phosphorus salts. *Asian Journal of Plant Sciences*, 5, 948-955. <<u>https://doi.org/10.3923/ajps.2006.948.955</u>>
- Bessho-Uehara, K., Furuta, T., Masuda, K., Yamada, S., Angeles-Shim, R.B., Ashikari, M. and Takashi, T. (2017). Construction of rice chromosome segment substitution lines harboring *Oryza barthii* genome and evaluation of yield-related traits. *Breeding Science*, 67, 408-415. <<u>https://doi.org/10.1270/jsbbs.17022</u>>

Bewley, J.D. and Nonogaki, H. (2017). Seed maturation and germination. In *Reference Module in Life Sciences*, Chapter 05092, Elsevier. https://doi.org/10.1016/B978-0-12-809633-8.05092-5

- Brooks, S., Athinuwat, D. and Chiangmai, P.N. (2020). Enhancing germination and seedling vigor of upland rice seed under salinity and water stresses by osmopriming. *Science & Technology Asia*, 25, 63-74. <<u>https://doi.org/10.14456/scitechasia.2020.21</u>>
- Cai, H.W. and Morishima, H. (2000). Genomic regions affecting seed shattering and seed dormancy in rice. *Theoretical and Applied Genetics*, 100, 840-846. <<u>https://doi.org/10.1007/s001220051360</u>>
- Chattopadhyay, B. and Kelley, K. (2016). Estimation of the coefficient of variation with minimum risk: a sequential method for minimizing sampling error and study cost. *Multivariate Behavioral Research*, **51**, 627-648. http://dx.doi.org/10.1080/00273171.2016.1203279
- Doku, G.D., Glover, M.K., Glover, E.K. and Dartey, K.P.A. (2016). The effect of the use of temperature on the breakage of dormancy and the subsequent performance of rice (*Oryza* spp.). *International Journal of Plant Science and Ecology*, 2, 1-9.
- Donohue, K., Dorn, L., Griffith, C., Kim, E., Aguilera, A., Polisetty, C.R. and Schmitt, J. (2005). Environmental and genetic influences on the germination of *Arabidopsis thaliana* in the field. *Evolution*, **59**, 740-757. <<u>https://doi.org/10.1111/j.0014-3820.2005.tb01750.x</u>>
- Fahimirad, S. and Ghorbanpour, M. (2018). Omics approaches in developing abiotic stress tolerance in rice (*Oryza sativa* L.). In *Advances in Rice Research for Abiotic Stress Tolerance* (Eds. M. Hasanuzzaman, M. Fujita, K. Nahar and J.K. Biswas), pp. 767-779, Woodhead Publishing. <<u>https://doi.org/10.1016/B978-0-12-814332-2.00038-1</u>>
- FAO (2021). Food outlook biannual report on global food markets. *Food Outlook*, November 2021. Rome. <<u>https://doi.org/10.4060/cb7491en</u>>
- FAOSTAT (2020). Food and Agriculture Organization of the United Nations. <<u>http://www.fao.org/faostat/</u>en/#data/QC Accessed 01/05/2020 to 04/015/2020>
- Finch-Savage, W.E. and Leubner-Metzger, G. (2006). Seed dormancy and the control of germination. *New Phytologist*, **171**, 501-523. https://doi.org/10.1111/j.1469-8137.2006.01787.x
- Finkelstein, R., Reeves, W., Ariizumi, T. and Steber, C. (2008). Molecular aspects of seed dormancy. Annual Reviews of Plant Biology, 59, 387-415. https://doi.org/10.1146/annurev.arplant.59.032607.092740>
- Futakuchi, K., Senthilkumar, K., Arouna, A., Vandamme, E., Diagne, M., Zhao, D., Manneh, B. and Saito, K. (2021). History and progress in genetic improvement for enhancing rice yield in sub-Saharan Africa. *Field Crops Research*, 267, article 108159. https://doi.org/10.1016/j.fcr.2021.108159>
- Ginestet, C. (2011). ggplot2: elegant graphics for data analysis. *Journal of the Royal Statistical Society:* Series A, **174**, 245-246. https://doi.org/10.1111/j.1467-985X.2010.00676 9.x>
- Graeber, K., Linkies, A., Steinbrecher, T., Mummenhoff, K., Tarkowská, D., Turečková, V., Ignatz, M., Sperber, K., Voegele, A., De Jong, H., Urbanová, T., Strnad, M. and Leubner-Metzger, G. (2014). Delay of germination 1 mediates a conserved coat-dormancy mechanism for the temperature- and gibberellin-dependent control of seed germination. *Proceedings of the National Academy of Sciences*, **111**, E3571-E3580. <<u>https://doi.org/10.1073/pnas.1403851111</u>>
- Grenier, E. (2007). Quelle est la « bonne » formule de l'écart-type? Revue Modulad, 102, 37.
- Gu, X.Y., Liu, T., Feng, J., Suttle, J.C. and Gibbons, J. (2010). The qSD12 underlying gene promotes abscisic acid accumulation in early developing seeds to induce primary dormancy in rice. *Plant Molecular Biology*, 73, 97-104. <<u>https://doi.org/10.1007/s11103-009-9555-1</u>>
- Gubler, F., Millar, A.A. and Jacobsen, J.V. (2005). Dormancy release, ABA and pre-harvest sprouting. *Current Opinion in Plant Biology*, **8**, 183-187. <<u>https://doi.org/10.1016/j.pbi.2005.01.011</u>>
- Hanumanthappa, D., Vasudevan, S.N., Maruthi, K., Maruthi, J.B. and Sebastian, A. (2016). Efficacy of dormancy breaking methods in paddy genotypes. *Journal of Applied and Natural Science*, 8, article 634. <<u>https://doi.org/10.31018/jans.v8i2.850</u>>
- Huang, X., Schmitt, J., Dorn, L., Griffith, C., Effgen, S., Takao, S., Koornneef, M. and Donohue, K. (2010). The earliest stages of adaptation in an experimental plant population: Strong selection on QTLs for seed dormancy. *Molecular Ecology*, **19**, 1335-1351. ">https://doi.org/10.1111/j.1365-294X.2010.04557.x>
- ISTA (2021). International Rules for Seed Testing, International Seed Testing Association, Bassersdorf, Switzerland.
- Kilian, B., Özkan, H., Pozzi, C. and Salamini, F. (2009). Domestication of the Triticeae in the Fertile Crescent. In *Plant Genetics and Genomics: Crops and Models* (Eds. G. Muehlbauer and C. Feuillet). 7, 81-119. Springer, New York, NY. <<u>https://doi.org/10.1007/978-0-387-77489-3_3</u>>
- Kumar, M., Rajpurohit, D., Basha, P.O., Bhalla, A., Randhawa, G.S. and Dhaliwal, H.S. (2009). Genetic Control of Seed Dormancy in Basmati Rice. *Madras Agriculture Journal*, 96, 7-12, 305-308.

- Kuya, N., Sun, J., Iijima, K., Venuprasad, R. and Yamamoto, T. (2019). Novel method for evaluation of anaerobic germination in rice and its application to diverse genetic collections. *Breeding Science*, 69, 633-639. <<u>https:// doi.org/10.1270/jsbbs.19003</u>>
- Lin, S.Y., Sasaki, T. and Yano, M. (1998). Mapping quantitative trait loci controlling seed dormancy and heading date in rice, *Oryza sativa* L., using backcross inbred lines. *Theoretical and Applied Genetics*, 96, 997-1003. <<u>https://doi.org/10.1007/s001220050831</u>>
- Lu, Q., Niu, X., Zhang, M., Wang, C., Xu, Q., Feng, Y., Yang, Y., Wang, S., Yuan, X., Yu, H., Wang, Y., Chen, X., Liang, X. and Wei, X. (2018). Genome-wide association study of seed dormancy and the genomic consequences of improvement footprints in rice (*Oryza sativa* L.). *Frontiers in Plant Science*, 8, 2213. https://doi.org/10.3389/fpls.2017.02213>
- Marzougui, S., Sugimoto, K., Yamanouchi, U., Shimono, M., Hoshino, T., Hori, K., Kobayashi, M., Ishiyama, K. and Yano, M. (2012). Mapping and characterization of seed dormancy QTLs using chromosome segment substitution lines in rice. *Theoretical and Applied Genetics*, **124**, 893-902 <<u>https://doi.org/10.1007/s00122-011-1753-y></u>
- Mendez del Villar, P. and Lançon, F. (2015). West African rice development: Beyond protectionism versus liberalization? *Global Food Security*, 5, 56-61. https://doi.org/10.1016/j.gfs.2014.11.001>
- Mizuno, Y., Yamanouchi, U., Hoshino, T., Nonoue, Y., Nagata, K., Fukuoka, S., Ando, T., Yano, M. and Sugimoto, K. (2018). Genetic dissection of pre-harvest sprouting resistance in an upland rice cultivar. *Breeding Science*, 68, 200-209. https://doi.org/10.1270/jsbbs.17062>
- Mutinda, Y.A., Muthomi, J.W., Kimani, J.M., Cheminigw'wa, G.N. and Olubayo, F.M. (2017). Viability and dormancy of rice seeds after storage and pre-treatment with dry heat and chemical agents. *Journal of Agricultural Science*, 9, 175-185. https://doi.org/10.5539/jas.v9n7p175
- Ndindeng, S.A., Candia, A., Mapiemfu, D.L., Rakotomalala, V., Danbaba, N., Kulwa, K., Houssou, P., Mohammed, S., Jarju, O.M., Coulibaly, S.S., Baidoo, E.A., Moreira, J. and Futakuchi, K. (2021). Valuation of rice postharvest losses in Sub-Saharan Africa and its mitigation strategies. *Rice Science*, 28, 212-216. <<u>https://doi.org/10.1016/j.rsci.2021.04.001</u>>
- Ndjiondjop, M.N., Gouda, A.C., Eizenga, G.C., Warburton, M.L., Kpeki, S.B., Wambugu, P.W., Gnikoua, K., Tia, D.D. and Bachabi, F. (2022). Genetic variation and population structure of *Oryza sativa* accessions in the AfricaRice collection and development of the AfricaRice *O. sativa* core collection. *Crop Science*, 63, 724-739. <<u>https://doi.org/10.1002/CSC2.20898</u>>
- Ndjiondjop, M.N., Semagn, K., Gouda, A.C., Kpeki, S.B., Tia, D.D., Sow, M., Goungoulou, A., Sie, M., Perrier, X., Ghesquiere, A. and Warburton, M.L. (2017). Genetic variation and population structure of *Oryza glaberrima* and development of a mini-core collection using DArTseq. *Frontiers in Plant Science*, 8, article 1748. <<u>https://doi.org/10.3389/fpls.2017.01748</u>>
- Nedunchezhiyan, V., Palanivel, M., Akhila Jabeen, P.A., Thangavel, P., Ramakrishnan, B., Velusamy, M., Muthusamy, S. and Edm, I.A. (2023). Effects of gibberellic acid on seed dormancy of black gram (Vigna mungo L.). Journal of Applied Biology and Biotechnology, 11, 256-259. <<u>https://doi.org/10.7324/JABB.</u> 2023.45506>
- Née, G., Kramer, K., Nakabayashi, K., Yuan, B., Xiang, Y., Miatton, E., Finkemeier, I. and Soppe, W.J.J. (2017a). DELAY of GERMINATION1 requires PP2C phosphatases of the ABA signalling pathway to control seed dormancy *Nature Communications*, 8, 72. https://doi.org/10.1038/s41467-017-00113-6>
- Née, G., Xiang, Y. and Soppe, W.J. (2017b). The release of dormancy, a wake-up call for seeds to germinate. *Current Opinion in Plant Biology*, 35, 8-14. <<u>https://doi.org/10.1016/j.pbi.2016.09.002</u>>
- Nelson, G.C., Valin, H., Sands, R.D., Havlík, P., Ahammad, H., Deryng, D., Elliott, J., Fujimori, S., Hasegawa, T., Heyhoe, E., Kyle, P., Von Lampe, M., Lotze-Campen, H., Mason D'Croz, D., Van Meijl, H., Van Der Mensbrugghe, D., Müller, C., Popp, A., Robertson, R., Robinson, S., Schmid, E., Schmitz, C., Tabeau, A. and Willenbockel, D. (2014). Climate change effects on agriculture: Economic responses to biophysical shocks. *Proceedings of the National Academy of sciences*, **111**, 3274-3279. <<u>https://doi.org/10.1073/</u>pnas.1222465110>
- Nguyen, T., Zhou, C., Zhang, T., Yu, J., Miao, R., Huang, Y., Zhu, X., Song, W., Liu, X., Mou, C., Lan, J., Liu, S., Tian, Y., Zhao, Z., Jiang, L. and Wan, J. (2019). Identification of QTL for seed dormancy from weedy rice and its application to elite rice cultivar 'Ninggeng 4'. *Molecular Breeding*, **39**, 1-12. <<u>https://doi.org/10.1007/ s11032-019-1031-2</u>>
- OECD/FAO (2019). OECD-FAO Agricultural Outlook 2019-2028, OECD Publishing, Paris/Food and Agriculture Organization of the United Nations, Rome. https://doi.org/10.1787/agr_outlook-2019-en>

- Olbana, T., Muchugi, A., Woldemariam, Y., Hay, F.R., Ndiwa, N. and Jones, C.S. (2023). Overcoming dormancy in *Desmanthus virgatus* seeds for improved viability monitoring protocols of genebank accessions. *Seed Science and Technology*, **51**, 421-434. https://doi.org/10.15258/sst.2023.51.3.12)
- Osinski, J. and Sylla, F. (2018). 2018 West Africa Rice Annual. Senegal, Global Agricultural Information Network, USDA Foreign Agricultural Service.
- Pandey, S., Byerlee, D., Dawe, D., Doberman, A., Mohanty, S., Rozelle, S. and Hardy, B. (2010). *Rice in the Global Economy: Strategic Research for Policy Issues and Food Security*, International Rice Research Institute, Philippines.
- Pandey, V. and Shukla, A. (2015). Acclimation and tolerance strategies of rice under drought stress. *Rice Science*, 22, 147-161. https://doi.org/10.1016/j.rsci.2015.04.001
- Pathak, H., Kumar, M., Molla, K.A. and Chakraborty, K. (2021). Abiotic stresses in rice production: Impacts and management. Oryza, 58, 103-125. https://doi.org/10.35709/ory.2021.58.spl.4>
- Ros, C., Bell, R.W. and White, P.F. (2000). Phosphorus seed coating and soaking for improving seedling growth of Oryza sativa (rice) cv. IR66. Seed Science and Technology, 28, 391-401.
- Saito, K., Dieng, I., Toure, A.A., Somado, E.A. and Wopereis, M.C.S. (2015). Rice yield growth analysis for 24 African countries over 1960-2012. *Global Food Security*, 5, 62-69. <<u>https://doi.org/10.1016/j.gfs.2014.10.006</u>>
- Sekiya, N., Oizumi, N., Kessy, T.T., Fimbo, K.M.J., Tomitaka, M., Katsura, K. and Araki, H. (2020). Importance of market-oriented research for rice production in Tanzania. A review. *Agronomy for Sustainable Development*, 40, 1-16. https://doi.org/10.1007/s13593-020-0611-1>
- Shalinee, D., Kumar, R., Chaudhari, S. and Pravina, P. (2019). Effect of GA₃ and IAA on dormancy and seedling growth of fresh sesame [*Sesamum indicum* (L.)] seeds. *Journal of Pharmacognosy and Phytochemistry*, 8, 561-565.
- Shi, J., Shi, J., Liang, W. and Zhang, D. (2021). Integrating GWAS and transcriptomics to identify genes involved in seed dormancy in rice. *Theoretical and Applied Genetics*, **134**, 3553-3562. <<u>https://doi.org/10.1007/ S00122-021-03911-1</u>>
- Shiratsuchi, H., Ohdaira, Y., Yamaguchi, H. and Fukuda, A. (2017). Breaking the dormancy of rice seeds with various dormancy levels using steam and high temperature treatments in a steam nursery cabinet. *Plant Production Science*, 20, 183-192. https://doi.org/10.1080/1343943X.2017.1289067>
- Shu, K., Liu, X.D., Xie, Q. and He, Z.H. (2016). Two faces of one seed: hormonal regulation of dormancy and germination. *Molecular Plant*, 9, 34-45. https://doi.org/10.1016/j.molp.2015.08.010>
- Sikirou, M., Saito, K., Achigan-Dako, E.G., Dramé, K.N., Ahanchédé, A. and Venuprasad, R. (2015). Genetic improvement of iron toxicity tolerance in rice-progress, challenges and prospects in West Africa. *Plant Production Science*, 18, 423-434. https://doi.org/10.1626/pps.18.423>

Statista (2023). statista.com

- R Development Core Team (2011). *R: A Language and Environment for Statistical Computing*, R Foundation for Statistical Computing, Vienna. <<u>http://www.R-project.org</u>>
- Timple, S.E., Hay, F.R., Mercado, M.F.O., Borromeo, T.H. and Sta. Cruz, P.C. (2018). Response of intact seeds of wild rice (Oryza) species to dry heat treatment and dormancy-breaking chemicals. *Seed Science and Technology*, **46**, 157-173. https://doi.org/10.15258/sst.2018.46.1.16
- Tung, L.D. and Serrano, E.P. (2011). Effects of warm water in breaking dormancy of rice seed. *Omonrice*, 18, 129-136.
- Veasey, E.A., Karasawa, M.G., Santos, P.P., Rosa, M.S., Mamani, E. and Oliveira, G.C.X. (2004). Variation in the loss of seed dormancy during after-ripening of wild and cultivated rice species. *Annals of Botany*, 94, 875-882. <<u>https://doi.org/10.1093/aob/mch215</u>>
- Waheed, A., Ahmad, H. and Abbasi, F.M. (2012). Different treatment of rice seed dormancy breaking, germination of both wild species and cultivated varieties (*Oryza sativa L.*). Journal of Materials and Environmental Science, 3, 551-560.
- Wan, J.M., Jiang, L., Tang, J.Y., Wang, C.M., Hou, M.Y., Jing, W. and Zhang, L.X. (2006). Genetic dissection of the seed dormancy trait in cultivated rice (*Oryza sativa* L.). *Plant Science*, **170**, 786-792. <<u>https://doi.org/10.1016/j.plantsci.2005.11.011</u>>
- Wu, Y. and Shen, Y.B. (2021). Sulfuric acid and gibberellic acid (GA₃) treatment combined with exposure to cold temperature modulates seed proteins during breaking of dormancy to germination in *Tilia miqueliana*. *The Protein Journal*, **40**, 940-954. https://doi.org/10.1007/s10930-021-10018-5

- Ye, C., Zhou, Q., Wu, X., Ji, G. and Li, Q.Q. (2019). Genome-wide alternative polyadenylation dynamics in response to biotic and abiotic stresses in rice. *Ecotoxicology and Environmental Safety*, **183**, article 109485. https://doi.org/10.1016/j.ecoenv.2019.109485
- Yuningsih, A.F.V. and Wahyuni, S. (2015). Effective methods for dormancy breaking of 15 new-improved rice varieties to enhance the validity of germination test. In *International Seminar on Promoting Local Resources* for Food and Health. Bengkulu, 12-13.
- Zenna, N., Senthilkumar, K. and Sie, M. (2017). Rice production in Africa. In *Rice Production Worldwide* (Eds. B.S. Chauhan, K. Jabran and G. Mahajan), pp. 117-135. <<u>https://doi.org/10.1007/978-3-319-47516-55></u>
- Zhang, C., Yuan, Z., Wang, Y., Sun, W., Tang, X., Sun, Y. and Yu, S. (2020). Genetic dissection of seed dormancy in rice (*Oryza sativa* L.) by using two mapping populations derived from common parents. *Rice*, **13**, 1-13. <<u>https://doi.org/10.1186/s12284-020-00413-4</u>>